



Gabriel A. Rabinovich

Profile

Dr. Gabriel Adrián Rabinovich obtained his BSc Biochemist degree in 1993 followed by his Ph.D. degree both from the School of Chemical Sciences (University of Córdoba). Currently, he is a Principal Investigator of the Argentinean National Research Council (CONICET), Head of the Laboratory of Immunopathology and Vicedirector of the Institute of Biology and Experimental Medicine, and Professor of Immunology at the Faculty of Exact and Natural Sciences (University of Buenos Aires). Also Dr. Rabinovich is Visiting Professor at the University of Maryland, Dana Farber Cancer Institute in Harvard Medical School and University of Miami. He serves as Editor of several journals including Glycobiology, Cell Death and Differentiation, Oncoimmunology, Immunology and Cell Biology and Emerging Infectious Diseases. Using an interdisciplinary approach, **Dr. Rabinovich's lab aims to identify different therapeutic targets, based on protein-glycan interactions, to differentially modulate immune-mediated disorders including cancer, autoimmunity and chronic inflammation.** For his scientific and professional achievements Dr. Rabinovich was awarded with the John Simon Guggenheim fellowship (USA), Third World Academy of Sciences (TWAS) Award in Medical Research (Italy), Bunge Born Award (Argentina), Bernardo Houssay Award (Argentina), and Cancer Research Institute Investigator Award (USA). He has published 145 articles in international leading journals including *Cancer Cell*, *Nature Immunology*, *Nature Medicine*, *Immunity*, *Journal of Experimental Medicine*, *Nature Rev Immunol*, *PNAS*, *J Immunol* and *Annu Rev Immunol*, has written several chapters in books, and presented eight patents of his work.

Regulatory circuits mediated by lectins and glycans in immune tolerance and inflammation

Gabriel A. Rabinovich, Ph.D.

Laboratorio de Immunopatología
Instituto de Biología y Medicina Experimental (IBYME)
Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET),
ARGENTINA

Homeostatic signals delivered in the form of immunosuppressive cytokines or inhibitory receptors are integrated into regulatory circuits that sustain peripheral tolerance mechanisms. These mechanisms serve to prevent collateral tissue damage resulting from over-exuberant immune responses to pathogens or seemingly-innocuous environmental stimuli¹⁾. Disruption of single pathways underlying these circuits leads to substantial inflammatory and autoimmune states. Conversely, their aberrant activation represents a significant hurdle for the development of anti-tumor immunity. Although under-appreciated for many years, exciting findings underscore the essential contribution of cell surface glycosylation and lectin-glycan signaling to these regulatory circuits operating in immune homeostasis. Indeed, endogenous glycan-binding proteins or lectins specifically decode glycan-containing information and convey this information into functional cellular responses²⁾.

Galectins are a family of soluble lectins characterized by a conserved carbohydrate recognition domain (CRD) that recognizes N- and O-glycans expressing the disaccharide N-acetylglucosamine (Galβ(1-4)-GlcNAc or LacNAc)³⁾. Secreted galectins, in contrast to cytokines or chemokines, do not have specific receptors, but can mediate cellular communication through recognition of a preferred set of cell surface glycoconjugates³⁾. **In our laboratory, we work at the interface of immunology and glycobiology using *in vitro* and *in vivo* approaches to understand the role of glycans and glycan-binding proteins in immune tolerance and inflammation.**

Research over the past decade revealed a prominent expression of galectin-1 in tumor cells and its association with malignant progression. These observations led us to investigate the role of galectin-1-glycan interactions in tumor biology.

By a combination of *in vitro* and *in vivo* experiments we identified a crucial role for galectin-1 in tumor-immune escape⁴⁾. We found that both human and murine melanoma cells secrete functional galectin-1, which substantially contributes to the immunosuppressive activity of these cells. Targeted inhibition of galectin-1 gene expression rendered mice resistant to tumor challenge and stimulated the generation of a tumor-specific Th1-type response⁴⁾. Supporting these findings, we found that Hodgkin lymphoma cells selectively overexpress galectin-1, favoring the secretion of Th2-type cytokines and blunting specific T-cell immunity⁵⁾. Notably, the observed role for galectin-1-glycan interactions in the establishment of immune privileged microenvironments was important not only in tumor settings but also at the fetomaternal interface⁶⁾. In an established model of stress-induced failing pregnancy, galectin-1 had a critical role in preventing fetal loss and restoring tolerance *in vivo*. Galectin-1-deficient (*Lgals1*^{-/-}) female mice showed higher rates of fetal loss compared to their wild-type counterparts in allogeneic, but not in syngeneic matings. Investigation of the mechanisms underlying these regulatory effects revealed the ability of galectin-1 to restore the Th1/Th2 cytokine balance and promote the expansion of IL-10-producing T_{reg} cells⁶⁾. These effects were recently confirmed in several autoimmune, inflammatory and tumor settings⁷⁾.

To understand the cellular mechanisms underlying these immunoregulatory effects, we performed *in vitro* and *in vivo* studies. We found a link between differential glycosylation of T helper cells, susceptibility to galectin-1-induced cell death and termination of the inflammatory response⁸⁾. While Th1- and Th17-differentiated cells expressed the repertoire of cell surface glycans that are critical for galectin-1 binding and cell death, Th2 cells were protected

from galectin-1 through differential α2,6 sialylation of cell surface glycoproteins⁸⁾. More recently, we investigated the relevance of galectin-glycan lattices within the dendritic cell (DC) compartment. Notably, DCs differentiated in a galectin-1-enriched microenvironment acquired a distinctive 'regulatory signature' characterized by abundant secretion of IL-27 and IL-10⁹⁾. When transferred *in vivo*, these DCs promoted antigen-specific T-cell tolerance, blunted Th1 and Th17 responses and halted autoimmune neuroinflammation through mechanisms involving DC-derived IL-27 and T cell-derived IL-10. Thus, using IL-27 receptor-deficient (*Il27ra*^{-/-}) and IL-10-deficient (*Il10*^{-/-}) mice, we have identified an immunoregulatory circuit linking galectin-1 signaling, IL-27-producing tolerogenic DCs and IL-10 secreting T_{reg} cells. Remarkably, galectin-1 expression increased during the peak and recovery phases of experimental autoimmune encephalomyelitis (EAE), and animal model of multiple sclerosis (MS), and was dramatically up-regulated by tolerogenic stimuli. Moreover, galectin-1-sufficient but not galectin-1-deficient DCs were able to restore tolerance and contribute to EAE recovery⁹⁾, suggesting a crucial role of endogenous galectin-1 in 'fine-tuning' the immunogenic function of DCs. Furthermore, we recently identified an essential role of galectin-1-glycan interactions in tempering microglia activation, brain inflammation and neurodegeneration with critical therapeutic implications in MS¹⁰⁾. We found that galectin-1 limits M1 microglia activation and neurodegeneration, by targeting the activation of p38MAPK- and

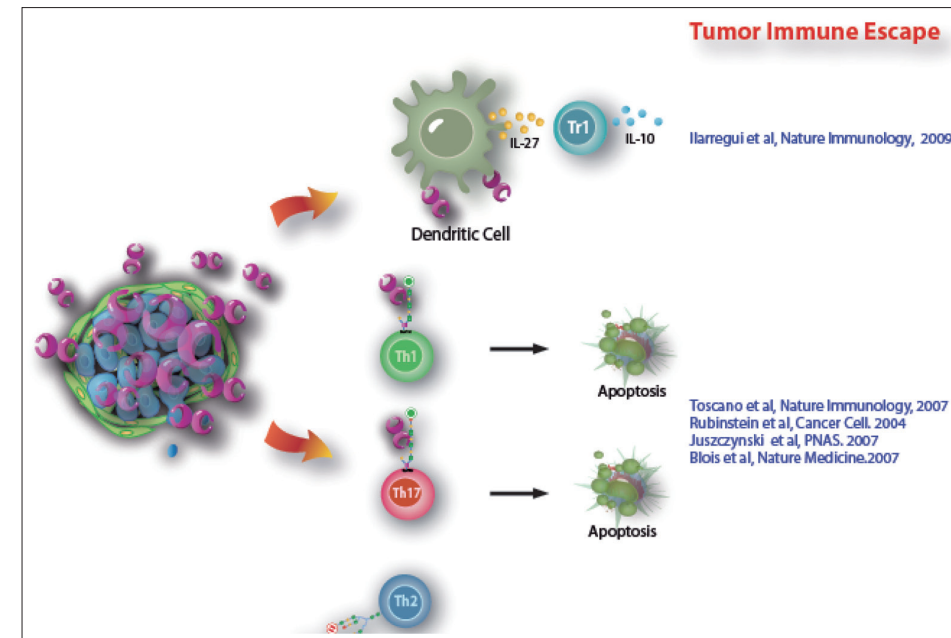


Figure 1. Tumour secreted galectin-1 promotes tumour immune escape through modulation of DCs differentiation and functionality and regulation of T cell homeostasis.

DCs differentiated in a galectin-1-enriched microenvironment acquire a tolerogenic phenotype characterized by abundant secretion of IL-27 and induction of IL-10 secreting T_{reg}. In addition, galectin-1 differentially regulates apoptosis of T cell populations. While Th1- and Th17-differentiated cells express the repertoire of cell surface glycans that are critical for galectin-1 binding and cell death, Th2 cells are protected from galectin-1 through differential α2,6 sialylation of cell surface glycoproteins.

CREB-dependent pathways and hierarchically controlling downstream pro-inflammatory mediators such as iNOS, TNF and CCL2. Mechanistically galectin-1 acted by specifically interacting on core 2 O-glycans on CD45 and retaining this glycoprotein on the surface of microglia cells, thus favoring its phosphatase activity and amplifying its inhibitory effect. Targeted deletion of galectin-1 resulted in pronounced inflammation-induced neurodegeneration in an animal model of multiple sclerosis. Moreover, adoptive transfer of astrocytes secreting galectin-1 or administration of recombinant galectin-1 rescued this phenotype

through mechanisms involving microglia de-activation¹⁰⁾. Thus, similar to the effects observed at the peripheral compartment, our results show that galectin-1-glycan interactions are key mediators of different cellular processes central to immune regulation in the central nervous system.

Taken together, our findings set the bases for the design of novel therapeutic strategies aimed at potentiating antitumor responses, preventing fetal loss and favoring the resolution of chronic inflammation and autoimmune disorders.

References

- 1) Bluestone JA, Thomson AW, Shevach EM, Weiner HL. What does the future hold for cell-based tolerogenic therapy? *Nat. Rev. Immunol.* (7): 650-654, 2007.
- 2) Rabinovich GA, Croci DO. Regulatory circuits mediated by lectin-glycan interactions in autoimmunity and cancer. *Immunity*. (36):322-335, 2012.
- 3) Rabinovich GA, Toscano MA. Turning 'sweet' on immunity: galectin-glycan interactions in immune tolerance and inflammation. *Nat. Rev. Immunol.* (9): 338-352, 2009.
- 4) Rubinstein N, Alvarez M, Zwirner NW, Toscano MA, Ilarregui JM, Bravo A, Mordoh J, Fainboim L, Podhajcer OL, Rabinovich GA. Targeted inhibition of galectin-1 gene expression in tumor cells results in heightened T cell-mediated rejection; A potential mechanism of tumor-immune privilege. *Cancer Cell* (5): 241-251, 2004.
- 5) Juszczynski P, Ouyang J, Monti S, Rodig SJ, Takeyama K, Abramson J, Chen W, Kutok JL, Rabinovich GA, Shipp MA. The API-1-dependent secretion of galectin-1 by Reed Sternberg cells fosters immune privilege in classical Hodgkin lymphoma. *Proc. Natl. Acad. Sci. U S A* (104): 13134-13139, 2007.
- 6) Blois SM, Ilarregui JM, Tometten M, Garcia M, Orsal AS, Cordo-Russo R, Toscano MA, Bianco GA, Kobelt P, Handjiski B, Tirado I, Markert UR, Klapp BF, Poirier F, Szekeres-Bartho J, Rabinovich GA, Arck PC. A pivotal role for galectin-1 in fetomaternal tolerance. *Nat. Med.* (13):1450-1457, 2007.
- 7) Cedeno-Laurent F, Watanabe R, Teague JE, Kupper TS, Clark RA, Dimitroff CJ. Galectin-1 inhibits the viability, proliferation, and Th1 cytokine production of nonmalignant T cells in patients with leukemic cutaneous T-cell lymphoma. *Blood*. (119):3534-3538, 2012.
- 8) Toscano MA, Bianco GA, Ilarregui JM, Croci DO, Correale J, Hernandez JD, Zwirner NW, Poirier F, Riley EM, Baum LG, Rabinovich GA. Differential glycosylation of TH1, TH2 and TH-17 effector cells selectively regulates susceptibility to cell death. *Nat. Immunol.* (8): 825-834, 2007.
- 9) Ilarregui JM, Croci DO, Bianco GA, Toscano MA, Salatino M, Vermeulen ME, Geffner JR, and Rabinovich GA. Tolerogenic signals delivered by dendritic cells to T cells through a galectin-1-driven immunoregulatory circuit involving interleukin 27 and interleukin 10. *Nat. Immunol.* (10): 981-991, 2009.
- 10) Starossom SC, Mascanfroni ID, Imitola J, Cao L, Raddassi K, Hernandez SF, Elyaman W, Basil R, Croci DO, Cerliani JP, Wang Y, Khoury SJ, Rabinovich GA. Galectin-1 deactivates classically-activated microglia and protects from inflammation-induced neurodegeneration. *Immunity*, 2012. in press.